Technology for their helpful discussion throughout this studies.

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Synthesis of 10-Oxo- β -rhodomycinone Derivatives

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Regiospecific total syntheses of (\pm) -11-deoxy-4-methoxy-10-oxo- β -rhodomycinone (21a) and (\pm) -11-deoxy-1-methoxy-10-oxo- β -rhodomycinone (21b) are described. 2-(2-Bromoethyl)-1,3-dioxane (6) was transformed to naphthalenone 12, which was condensed with (phenylsulfonyl)-isobenzofuranone 13 to afford 7,8-dihydro-9-ethyl-6-hydroxy-4-methoxynaphthacen-5,12-dione (15). Epoxide 16 prepared from olefinic compound 15, reacted with HF/Pyr (7:3) to give 17. Dihydroxylation of 17 with t-BuOK/P(OMe)₃/O₂, selective cis-diol protection of mixed compounds 18 with phenylboronic acid in toluene, separation of cis-boronate 19 and transdiol 20 by column chromatography on silica gel, and cleavage of the boronate group of 19 with 2-methylpentane-2,4-diol in acetic acid completed the construction of 21.

Introduction

Rhodomycins in anthracycline series were first discovered by Krassilnikov and Koreniakov.¹ Among these, rhodomycin and isorhodomycin family produced by *streptomyces purpurascens* were the first anthracycline compounds whose structures were elucidated by Brockmann and coworkers.²

Rhodomycins like daunomycin and aklavin are well-known, clinically useful anticancer chemotherapeutic agents against acute leukemia and human cancer, as well as various experimental tumors. Since then, anthracyclines have been the objects of intensive clinical tests and synthetic studies for the last decade because of their strong antineoplastic activities.³ Numerous synthetic approaches have been

Figure 1.

developed and a number of their derivatives have been synthesized.4

All rhodomycins have a hydroxyl group at C-11 position of the aglycone, and β -isorhodomycins (2) possess an additional hydroxyl group at C-1 position unlike β -rhodomycins (1). Cardiotoxicity studies by Tone and coworkers⁵ have shown that the toxicity was reduced when aclacinomycin derivatives without hydroxyl group at C-11 position were used. Moreover, 11-deoxydaunorubicin isolated by Arcamone et al.6 was found to have higher anticancer activity and less cardiotoxicity than daunorubicin having a hydroxyl group at C-11 position.

In a recent publication,7 we reported the synthesis of novel 9-fluoroanthracycline derivatives and their isomeric glycosides. The first synthesized 9-fluoroanthracycline derivative 3 showed an improved anti-proliferative effect to Hepatoma cells (Hep 3B). Here, we report the synthesis of aglycones (\pm) -21a and (\pm) -21b for the synthesis of anthracycline derivatives 4 and 5. The fluorine atom at C-9 position of 3 was replaced by an hydorxyl group.

Results and Discussion

The anthracyclines containing the tetracyclic polynuclear aromatic ring system in the aglycone have at least one sugar residue glycosidically linked4 to the C-7 hydroxyl group. The regiospecific convergent syntheses of aglycones were accomplished through condensation of phthalide sulfone derivatives with α,β -unsaturated carbonyl system using the Michael type condensation reaction.8 The key element of the synthesis was the use of naphthalenone 12,8a which served as a synthon for the construction of tetracyclic ring system by condensing with phthalide sulfone in one step. In the earlier work, 8a Rho reported the synthesis of 1(4H)-naphthalenone 12 through five steps involving Grignard reaction, acetylation, and elimination in the presence of palladium acetate as a catalyst. However, the Rho's route to prepare naphthalenone 12 took a long step. We have developed a short and efficient route for the preparation of the intermediate 9 in multigram quantities via direct Wittig reaction,9 and have employed the method to accomplish the synthesis of 10-oxo- β -rhodomycinones. Unlike the result from Grignard reaction (E-isomer only), the diene 9 synthesis through Wittig reaction gave a mixture of E- and Z-isomer (1:1). Without further purification, Diels-Alder cycloaddition of 9 with methyl vinyl ketone 10 (neat, sealed tube, 150 °C) regiospecifically furnished the cyclohexene 11 as a mixture of cis and trans isomers. The desired naphthalenone 12 was successfully synthesized in 80% yield by the hydrolysis of acetal group

Scheme 1. (a) PPh₃/toluene, reflux, (b) n-BuLi/THF, acrolein (8), (c) methyl vinyl ketone (10) in seal tube, 150 °C, (d) 3 N

using 3 N aqueous HClO₄.

Naphthalenone 12 was condensed with phthalide sulfone 13 which had been converted to an anion with t-BuOLi at -78 °C to afford naphthacene 15 in 83-85% yield after oxygen bubbling in DMF, and then the double bond of 15 was subjected to epoxidation with m-CPBA to produce epoxide 16.10 From the previous studies on the syntheses of 9-fluoro-10-hydroxynaphthacene and 10-oxo compound 17,7,11 we found that the fluorinated compound was obtained as a major product when the epoxy compound 16 was treated with HF/pyridine (7:3) at 0 °C for 5 min. On the other hand, when the same reaction was performed at room temperature for 8 hr, the epoxide 16 was transformed to 10oxo compound 17 in 70% yield. It appears that the epoxide 16 in HF/pyridine (7:3) generated the fluorinated compound within 5 min and then dehydrofluorination took place due to the pyridine base.

Now, the two hydroxyl groups have to be introduced at C-7 and C-9 position as cis form to obtain the final products 21. Hydroxylation of 10-oxo-compound 17 at C-7 and C-9 position was achieved by Swenton's method¹² (O₂/t-BuOK/P(OCH₃)₂/DMF). 9-Hydroxyl compound and 7,9dihydroxy compound 18 were obtained in 12% and in 53% yield respectively. Tanaka et al.13 reported that the amount of t-BuOK was a critical factor to give the best result of cis/ trans formation. After several trials, we obtained the 7.9dihydroxy compound 18 in the best yield when using 4 equivalent of t-BuOK. The major product of the reaction

Scheme 2. (a) t-BuOLi/THF, -78 °C, rt. reflux, (b) O₂/DMF, (c) m-CPBA/CH₂Cl₂, rt, 5 hr., (d) HF/Pyr (7:3)/THF, 0 °C (5 min), rt (8 hr), (e) t-BuOK, P(OMe)₃, O₂/DMF, -15 °C, (f) PhB (OH)₂, p-TsOH/toluene, (g) 2-methyl-2,4-pentanediol, AcOH/ CH₂Cl₂/acetone.

was cis compound: the cis/trans ratio was 9:1, which was determined by ¹H NMR spectrum analysis. The selective cisdiol protection of the isomers 18 with phenylboronic acid in dry toluene gave the benzene boronate (\pm) -19 and the unreacted trans-diol (\pm) -20.12 After purification by chromatography, the isolated cis-boronate (\pm) -19 was easily converted to cis-diol (\pm)-21 using 2-methylpentane-2, 4-diol in acetic acid. 14 The structures of 20 and 21 having a half-chair form⁶ were analyzed based on the ¹H NMR spectra. The protons at C-7 and C-8 exhibited different scalar coupling patterns for cis-21a ($J_{7e,8a}$ =3.91 Hz, $J_{7e,8e}$ = 1.96 Hz) and trans-20a ($J_{7a,8a}$ =8.79 Hz, $J_{7a,8c}$ =5.86 Hz), which were similar to the results of Keay.¹⁵ 1-Methoxy compound 21b was readily prepared by the same procedure for the synthesis of 4-methoxy compound 21a, and all the reactivities for the synthesis of 21b were similar to those of **21a. 21b** ($J_{7e,8a}$ =4.40 Hz, $J_{7e,8e}$ =3.01 Hz) and **20b** ($J_{7a,8a}$ =9.28 Hz, $J_{7a.8e}$ =5.86 Hz) were also identified as cis and trans isomer, respectively, by comparing the coupling constants with those reported in the literature.15

In conclusion, we developted an efficient method for the synthesis of the title compound (\pm) -21, which was the precursor for the synthesis of the glycoside as a potential anticancer drug. The glycoside containing N-acetyl-D-glucosamine or L-fucose can be readily prepared in a few steps. The *in vitro* antitumor activity of the glycosides against adrimycin will be reported elsewhere.

Experimental

All reactions were carried out under nitrogen atmosphere with oven-dried glassware. All solvents were purified by distillation and dried, if necessary, prior to use. ¹H and ¹³C spectra were obtained on a JEOL JNM EX-400 spectrometer. Chemical shifts were internally referenced to TMS for ¹H or to the solvent signals for ¹³C. Mass spectra were obtained on a JEOL JMX-DX 300 spectrometer (EI and FAB+) and on a VG QUATTRO spectrometer (ESI). Melting points were obtained on a Buchi 510 melting point apparatus and were uncorrected. Products were purified by flash column chromatography on silica gel (60-200 mesh), and HPLC was carried out on a Waters 4000 instrument having a Waters PDA UV spectrophotometer and a Waters 410 differential refractometer.

2-(4-Methylene-2-hexenyl)-1,3-dioxane (9). 2-(2-bromoethyl)-1,3-dioxane **6** (4.91 g, 25.17 mmol) and triphenylphosphine (6.60 g, 25.17 mmol) in tolune (100 mL) were heated at reflux under dry nitrogen for 2 days. The cooled product was separated by filtration, washed well with dry ether, and dried under vacuum to afford phosphonium salt **7** (10.50 g, 91.2%).

7 (3.13 g, 6.85 mmol) was suspended in 20 mL of tetrahydrofuran under dry nitrogen and treated with a solution of n-buthyllithium in hexane (1.2 molar equiv., 5.9 mL of a 1.4 M solution) at -20 °C. After 15 min, the nearly clear, red solution was cooled to -78 °C, and acrolein 8 (0.71 mL, 6.16 mmol) was added, discoloring the solution. The solution was then warmed to room temperature. The solvent was evaporated, and the oil was dissolved in methylene chloride. This solution was rinsed with 5% aqueous sodium sulfate solution and concentrated to dienes 9 (0.99 g, 88.5%)

as liquid. The physical and spectroscopic data of **9** are in accordance with the literature.^{8a}

1-Acetyl-2-[(1,3-dioxa-2-cyclohexyl)methyl]-4-ethylcyclohex-3-ene (11). Diene 9 (4.5 g, 24.69 mmol) was treated with methyl vinyl ketone 10 (3.1 mL, 37.04 mmol) in a sealed tube according to our earlier procedure to give adduct 11 (5.98 g, 96%). The physical and spectroscopic data are in accordance with the literature.^{8a}

6-Ethyl-4a,7,8,8a-tetrahydro-1(4H)-naphthalenone (12). Adduct **11** (5.0 g, 19.81 mmol) was hydrolyzed with 3 N HClO₄ (30 mL) according to our earlier procedure to give naphthalenone **12** (2.79 g, 80%). Less polar isomer: ¹H NMR (CDCl₃) δ 6.98 (ddd, J=10.2, 5.5, 2.4 Hz, 1H), 6.05 (ddd, J=10.2, 2.4, 1.9 Hz, 1H), 5.22 (dtt, J=3.4, 1.9, 1.5 Hz, 1H), 2.30-2.55 (m, 3H), 2.0-2.3 (m, 4H), 1.95 (q, J=7.0 Hz, 1H), 1.36 (m, 1H), 1.02 (t, J=7.0 Hz, 3H). More polar isomer: 6.86 (dt, J=10.2, 4.4 Hz, 1H), 5.99 (dt, J=10.2, 1.9 Hz, 1H), 5.30 (dtt, J=5.9, 2.4, 1.5 Hz, 1H), 2.8 (m, 1H), 2.45-2.63 (m, 2H), 2.15-2.25 (m, 1H), 2.0-2.10 (m, 3H), 1.95 (q, J=7.0 Hz, 2H), 1.68 (m, 1H), 0.97 (t, J=7.0 Hz, 3H).

7,8-Dihydro-9-ethyl-6-hydroxy-4-methoxy-naphthacen-5,12-dione (15a). 7,8-Dihydro-9-ethyl-6-hydroxy-1-methoxynaphthacen-5,12-dione (15b).

To cold (-78 °C) magnetically stirred solution of lithium t-Butoxide, prepared from t-butyl alcohol (6.13 mL, 64.06 mmol) and n-BuLi (45.76 mL, of 1.4 M solution, 64.06 mmol) in dry THF (70 mL) was added the sulfone 13a (7.22 g, 23.73 mmol) in THF (50 mL), and the mixture was stirred for 30 min at -78 °C. Solution of 12 (3.76 g, 21.35 mmol) in THF (20 mL) was added by syringe, and the mixture was stirred at -78 °C for 2 h. The cooling bath was removed, and the reaction was stirred at r.t. for 3 h and then heated at reflux for 20 min. The mixture was cooled to 0 °C and acidified with 2 N HCl. The THF was evaporated under reduced pressure, and the aqueous mixture was extracted with methylene chloride. The combined methylene chloride extracts were washed successively with H₂O, brine, dried MgSO₄, filtered, and evaporated at reduced pressure to give crude naphthacenone 14a (6.44 g, 89.2%). Oxygen was bubbled through a heated (100 °C) solution of 14a (2.7 g, 7.98 mmol) in DMF (50 mL) for 6 h. The solution was cooled in an ice-water bath. Addition of H₂O to the solution precipitated 15a as orange crystals, which were collected by filtration, washed with H₂O, and dried to give 2.23 g (83.6%) of pure **15a** with mp 156-158 °C.

15a: ¹H NMR (CDCl₃) δ 13.28 (s, 1H), 7.95 (dd, J=8.0, 1.9 Hz, 1H), 7.71 (t, J=8.0 Hz, 1H), 7.44 (s, 1H), 7.34 (dd, J=8.0, 1.0 Hz, 1H), 6.29 (s, 1H), 4.07 (s, 3H), 2.94 (t, J=8.8 Hz, 2H), 2.34 (t, J=8.8 Hz, 2H), 2.27 (q, J=7.6 Hz, 2H), 1.15 (t, J=7.6 Hz, 3H); MS m/z 334 (M⁺).

By the above procedure, 15b was obtained in a yield of 65% over two steps from 13b.

15b: mp 168-170 °C; ¹H NMR (CDCl₃); δ 12.78 (s, 1H), 7.93 (d, J=7.8 Hz, 1H, ArH), 7.68 (t, J=8.1 Hz, 1H, ArH), 7.41 (s, 1H, ArH), 7.38 (d, J=8.3 Hz, 1H, ArH), 6.29 (s, 1H, =CH), 5.03 (s, 3H, OMe), 2.90 (t, J=8.1 Hz, 2H, CH₂), 2.34-2.27 (m, 4H), 1.16 (t, J=7.31 Hz, 3H, CH₃); MS (m/z) 334 (M⁺).

9,10-Epoxy-9-ethyl-6-hydroxy-4-methoxy-7,8,9, 10-tetrahydronaphthacen-5,12-dione (16a). 9,10-

Epoxy-9-ethyl-6-hydroxy-1-methoxy-7,8,9,10-tetrahydronaphthacen-5,12-dione (16b). A mixture of naphthacene 15a (2.45 g, 7.33 mmol) and *m*-CPBA (70%, 2.71 g, 10.99 mmol) was dissolved in 50 mL of methylene chloride and stirred at r.t. for 5 h. The reaction mixture was diluted with 50 mL of methylene chloride and washed successively with 10% aqueous sodium carbonate solution and saturated brine, dried over Na₂SO₄. The residue was chromatographed on silica gel (CH₂Cl₂/EtOAc=95:5) to give 2.40 g (93.3%) of 16a with mp 208-210 °C. 16a: ¹H NMR (CDCl₃) δ 13.25 (s, 1H), 7.96 (dd, J=7.8, 0.9 Hz, 1H), 7.80 (s, 1H), 7.73 (t, J=7.8 Hz, 1H), 7.34 (d, J=7.8 Hz, 1H), 4.07 (s, 3H), 3.73 (s, 1H), 3.08 (m, 2H), 2.48 (m, 2H), 1.88 (q, J=7.9 Hz, 2H), 1.06 (t, J=7.9 Hz, 3H); MS m/z 350 (M⁺).

By the above procedure, 16b was obtained from 15b in 92.1% yield with mp 184-186 °C.

16b: ¹H NMR (CDCl₃) δ 12.67 (s, 1H, OH), 7.85 (d, J= 7.8 Hz, 1H, ArH), 7.68 (t, J=8.1 Hz, 1H, ArH), 7.41 (s, 1H, ArH), 7.39 (d, J=8.3 Hz, 1H, ArH), 6.29 (s,1H), 4.03 (s, 3H, OMe), 2.90 (t, J=8.5, 2H, CH₂), 2.34-2.27 (m, 4H), 1.16 (t, J=7.3 Hz, 3H, CH₃); MS (m/z) 350 (M⁺).

9-Ethyl-6-hydroxy-4-methoxy-7,8,9,10-tetrahydronaphthacen-5,10,12-trione (17a). 9-Ethyl-6-hydroxy-1-methoxy-7,8,9,10-tetrahydronaphthacen-5,10,12trione (17b). Using polyethylene equipment and under strict exclusion of moisture, to the cooled (0 °C) solution of HF/Pyr (7:3, 54 mL) was added the epoxide **16a** (1.70 g, 4.85 mmol) in THF (50 mL) in one portion, and the mixture was stirred for 5 min. The cooling bath was removed and the mixture was stirred for 8 h at r.t. The reaction mixture was poured into ice water (100 mL) and extracted with CH₂Cl₂. The combined organic phases were evaporated to dryness and the residue was purified by flash chromatography on silica gel (CH₂Cl₂/EtOAc, 2% → CH₂Cl₂ /EtOAc, 10%) to give 17a (1.44 g, 84.7%) with mp 208-210 °C as light yellow powder. 17a: ¹H NMR (400 MHz, CDCl₃): δ 13.37 (s, 1H, OH), 8.33 (s, 1H, ArH), 7.96 (d, 1H, J=8.1 Hz, ArH), 7.77 (t, 1H, J=8.1 Hz, ArH), 7.37 (d, 1H, J=8.1 Hz, ArH), 4.08 (s, 3H, OMe), 3.22 (dt, 1H, J=18.6, 4.9 Hz, $C_{7eg}H$), 2.89 (ddd, J=18.6, 9.7, 4.9 Hz, 1H, $C_{7ex}H$), 2.32 (ddd, 1H, J=13.6, 9.7, 4.7 Hz, 1H, C_{8ea}H), 2.02-1.90 (m, 2H), 1.63-1.53 (m, 1H, $C_{13}H$), 1.59-1.46 (m, 1H, $C_{9ax}H$), 1.03 (t, 3H, J=7.3 Hz, 3H, C_{14} H); ¹³C NMR (150 MHz, CDCl₃): δ 198.91, 188.91, 181.88, 161.01, 160.69, 140.28, 137.34, 136.17, 135.95, 130.31, 120.71, 120.30, 119.21, 118.30, 116.93, 56.67, 48.41, 26.34, 22.14, 22.03, 11.34; MS: m/z=350 (M⁺, 83%), 322 (100), 304 (32), 266 (27).

By the above procedure, 17b was obtained from 16b in 82.1% yield with mp 200-202 °C.

17b: ¹H NMR (400 MHz, CDCl₃) δ 12.74 (s, 1H, OH), 8.27 (s, 1H, ArH), 7.87 (d, 1H, J=7.8 Hz, ArH), 7.66 (t, 1H, J=8.1 Hz, ArH), 7.30 (d, 1H, J=8.3 Hz, ArH), 4.00 (s, 3H, OMe), 3.11 (dt, 1H, J=18.1, 4.4 Hz, 1H, C_{7eq} H), 2.88 (ddd, 1H, J=18.1, 9.7, 4.9 Hz, 1H, C_{9ax} H), 2.30 (m, 1H, C_{9ax} H), 2.25 (ddd, J=13.2, 4.9, 4.4 Hz, 1H, C_{8eq} H), 1.96-1.80 (m, 1H), 1.56-1.46 (m, 1H, C_{13} H), 0.95 (t, J=7.3 Hz, 3H, C_{14} H); ¹³C NMR (300 MHz, CDCl₃): δ 198.91, 188.84, 180.94, 160.61, 160.12, 138.51, 138.11, 135.23, 135.06, 129.79, 121.71, 119.52, 118.83, 116.37, 56.61, 48.50, 29.67, 26.35, 22.13, 21.86, 11.36; MS: m/z=350 (M*, 83%), 322 (100), 304 (32), 266 (27).

 (\pm) -11-Deoxy-4-methoxy-10-oxo- β -rhodomycinone (20a). (\pm)-4,11-Dideoxy-1-methoxy-10-oxoβ-rhodomycinone (20b). Oxygen was bubbled to the cold (-15 °C) magnetically stirred solution of t-BuOK (95%, 1.43 g, 12.10 mmol) and trimethyl phosphite (0.79 mL, 6.66 mmol) in dry DMF (30 mL) for 15 min. Solution of 17a (1.06 g, 3.03 mmol) in DMF (10 mL) was added, and the mixture was carefully maintained between - 15 and - 10 °C. After the color of the reaction mixture changed from yellow to dark blue, the reaction was quenched by the addition of 20 mL of water. The reaction mixture was stirred for 3 h at ambient temperature, the color of the mixture changing from dark blue to red-brown. The solvent was removed, and the resulting slurry was dissolved in CH₂Cl₂. The organic layer was then washed with water, filtered, and concentrated, and the trimethyl phosphate was removed under vacuum. The residue was chromatographed on silica gel (CH₂Cl₂/EtOAc 2%, \rightarrow CH₂Cl₂/EtOAc 50%) to give 18a (0.98 g, 85.1%).

The mixture of **18a** (0.72 g, 1.88 mmol), phenylboronic acid (97%, 0.31 g, 2.45 mmol) and anhydrous p-toluene-sulfonic acid (51 mg, 0.30 mmol) in dry toluene (50 mL) was stirred at r.t. for 13 h. The reaction mixture was quenched with saturated aqueous NaHCO₃ and extracted with CH₂Cl₂. The extract was washed with H₂O, dried, and concentrated *in vacuo* to give crude **19a**. Recrystallization from CHCl₃/hexane afforded **19a** (0.62 g, 70.8%) as red crystals.

The mixture of 19a (0.54 g, 1.15 mmol), 2-methyl-2,4pentanediol (1.47 mL, 11.53 mmol), AcOH (0.15 mL), CH₂Cl₂ (10 mL) and acetone (10 mL) was stirred at r.t. for 13 h. The reaction mixture was poured into a mixture of CH₂Cl₂ (30 mL) and saturated aqueous NaHCO₃ (15 mL). The organic layer was separated, washed with water, dried, and concentrated in vacuo. The residue was purified by flash chromatography on silica gel (CH₂Cl₂/EtOAc 1% → $CH_2Cl_2/EtOAc$ 5%), to give **21a** (0.38 g, 86.5%) with mp 175 °C as dark yellow powder. 21a: ¹H NMR (CDCl₃+ CD₃OD) δ 13.78 (s, 1H, OH), 8.47 (s, 1H, ArH), 8.02 (dd, J=7.8, 1.0 Hz, 1H, ArH), 7.82 (t, J=8.3 Hz, 1H, ArH), 7.41 (dd, J=8.8, 1.0 Hz, 1H, ArH), 5.40 (ddd, J=5.9, 3.9, 2.0 Hz, 1H, $C_{7ca}H$), 4.10 (s, 3H, OMe), 3.71 (s, 1H, C_7OH), 2.99 (dd, J=15.6, 3.9 Hz, 1H, C_{8eq} H), 2.84 (dd, J=15.6, 5.9 Hz, 1H, $C_{8ax}H$), 2.23 (dq, J=15.1, 7.3 Hz, 1H, $C_{13}H$), 1.16 (t, J=7.3 Hz, 3H, $C_{14}H$); ¹³C NMR (CDCl₃) δ 189.75, 188.77, 181.49, 161.25, 161.12, 136.72, 136.56, 135.63, 133.99, 132.40, 120.53, 120.39, 119.22, 118.42, 117.90, 64.27, 61.10, 56.78, 40.89, 32.28, 9.82; MS (m/z) 282 (M+).

20a: ¹H NMR (CDCl₃+CD₃OD) δ 14.06 (s, 1H, OH), 8.50 (s, 1H, ArH), 8.01 (dd, J=7.8, 1.0 Hz, 1H, ArH), 7.82 (t, J= 8.3 Hz, 1H, ArH), 7.41 (dd, J=8.8 Hz, 1H, ArH), 5.45 (ddd, J=8.8, 5.9, 1.9 Hz, 1H, C_{7αx}H), 4.54 (d, J=1.9 Hz, 1H, C₇OH), 4.10 (s, 3H, OMe), 3.01 (dd, J=14.7, 5.9 Hz, 1H, C_{8eq}H), 2.41 (dq, J=16.6, 7.3 Hz, 1H, C₁₃H), 2.28-2.22 (m, 2H, C_{8ax}H), 1.15 (t, J=7.3 Hz, 3H, C₁₄H); ¹³C NMR (CDCl₃) δ 189.10, 188.85, 181.36, 161.23, 160.84, 138.44, 136.80, 135.60, 134.21, 131.94, 120.53, 120.28, 118.89, 118.48, 118.34, 65.09, 56.75, 40.78, 32.16, 9.71; MS (m/z) 282 (M⁺).

By the above procedure, **21b** was obtained in a yield of 82.1% over three steps from **17b**.

21b: mp 155-158 °C; ¹H NMR (CDCl₃) δ 13.03 (s, 1H,

OH), 8.41 (s, 1H, ArH), 8.01 (dd, J=7.8, 1.0 Hz, 1H, ArH), 7.78 (t, J=8.3 Hz, 1H, ArH), 7.42 (d, J=8.8, 1.0 Hz, 1H, ArH), 5.06 (ddd, J=4.4, 3.0, 1.9 Hz, 1H, $C_{\tau eq}$ H), 4.07 (s, 3H, OMe), 2.96 (s, 1H, C_{τ} OH), 2.53 (dd, J=14.7, 3.0 Hz, 1H, $C_{\tau eq}$ H), 2.24 (dd, J=14.7, 4.4 Hz, 1H, $C_{\tau eq}$ H), 1.81-1.76 (m, 2H, $C_{\tau eq}$ H), 0.92 (t, J=7.1 Hz, 3H, $C_{\tau eq}$ H); $^{\tau eq}$ C NMR (CDCl₃) δ 199.91, 188.70, 180.40, 160.65, 160.42, 136.01, 135.56, 134.21, 133.03, 132.88, 121.79, 119.79, 118.92, 118.02, 117.84, 70.55, 63.52, 56.69, 40.18, 7.29; MS (m/z), 282 (M $^{+}$).

20b: ¹H NMR (CDCl₃) δ 13.36 (s, 1H, OH), 8.38 (s, 1H, ArH), 8.00 (dd, J=7.8, 1.0 Hz, 1H, ArH), 7.80 (t, J=8.0 Hz, 1H, ArH), 7.43 (dd, J=8.1, 1.0 Hz, 1H, ArH), 5.37 (ddd, J=9.3, 6.1, 3.4 Hz, 1H, C_{7ax} H), 4.07 (s, 3H, OMe), 3.66 (d, J=3.4 Hz, 1H, C_{7} OH), 2.79 (dd, J=13.7, 6.1 Hz, 1H, C_{8eq} H), 2.24 (dd, J=13.7, 9.3 Hz, 1H, C_{8ax} H), 1.76-1.56 (m, 2H, C_{13} H), 0.92 (t, J=7.3, Hz, 3H, C_{14} H); ¹³C NMR (CDCl₃) δ 199.45, 189.17, 180.15, 160.81, 160.31, 137.47, 135.51, 135.19, 134.84, 132.56, 121.37, 119.70, 119.47, 119.35, 117.89, 76.19, 63.63, 40.28, 29.82, 7.24; MS (m/z) 282 (M⁺).

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Rapid Energy Transfer Mechanism of F Electronic Excitation to the Vibration of Randomly Distributed OH in KCl

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The nature of F electronic excitation energy transfer to OH vibrational levels in KCl crystals is the exchange interaction, although the transfer process exhibits three temporally distinguishable components depending on the distance between excited F center and OH. The critical distance as well as rate of the major energy transfer process in randomly distributed samples increases rapidly as OH librational motions become active with temperature rise. The excited state character introduced into the OH ground electronic state by perturbation is essential for the exchange interaction. The perturbation is brought about by the expanded electron cloud of excited F center for OH associated to F center, whereas by librations and lattice vibrations perpendicular to the bond axis for isolated OH. F excitation quenching efficiency by OH is dependent on the variation of the critical distance rather than the rate as the rate is much faster than the normal F bleach recovery rate.

Introduction

This is the final paper concluding a series of studies 1-4 on

the bleach recovery kinetics of F centers in alkali halides. F absorption bleach in OH⁻ doped crystals recovers with four temporally distinguishable components designated as super-