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Gas Phase Investigation of $[(Cu^{2+}, Ni^{2+}--Gly-Gly-His) - 3H^{+}]^{-1}$ Complex by Electrospray Ionization MS/MS and MS/MS/MS

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Mass spectrometry (MS) is a very useful means by which to study the interactions of metal cation-biomolecule complexes in the gas phase. ^{1,2} The analysis of the fragmentation patterns of metal cationized peptides produced under electrospray ionization (ESI)-MS can provide complementary information for peptide sequencing when the fragmentation of the protonated peptide is insufficient. ^{3,4} The specific interactions in metal ion-peptide systems have been studied to develop practical sensors for the detection and quantification of metal ions. ⁵⁻⁷

Complexes of transition metal cations and peptides (transition metal²⁺--- peptide)²⁺ have been studied by many research groups.^{8,9} However, investigations regarding the [(Metal²⁺--- peptide) – 3H⁺]⁻¹ anion complex have not been conducted systematically using MS.^{7,10,11} The copper and nickel binding peptide Gly-Gly-His has been investigated in aqueous solution because the peptide Gly-Gly-His mimics the form of the specific Cu²⁺, Ni²⁺-transport active site of human serum albumin.^{12,13}

Theoretical studies concerning metal-oligopeptide structure and metal-ligand coordination geometry have also been performed through molecular dynamics simulations and *ab initio* calculations. ¹⁴⁻¹⁷ Structures, molecular orbital and stabilization energies of metal-oligopeptides are reported by the research groups.

In this study, our attention was focused on the interaction between the oligopeptide of three amino acid residues Gly-Gly-His and metal ions (Cu²⁺, Ni²⁺) in the gas phase. The interaction between the Gly-Gly-His and metal ions was studied by ESI-MS in negative mode. The fragmentation pattern of the [(Cu²⁺, Ni²⁺---Gly-Gly-His) – 3H⁺]⁻¹ anion complex was analyzed by MS/MS and MS/MS/MS spectra.

Experimental Section

The gas phase [(Metal²⁺---Gly-Gly-His) – 3H⁺]⁻¹ anion complex was produced by an electrospray ionization source. The experimental MS, MS/MS and MS/MS/MS data for fragmentation pattern analysis were obtained using a Thermo Finnigan LTQ mass spectrometer (Thermo Electron Corp., San Jose, CA, USA). This mass spectrometer is a linear ion trap mass spectrometer equipped with an atmospheric pressure-ionization source.

LTQ conditions. All spectra were acquired in negative

ion mode over a range of m/z 100-400 by averaging 40 scans. The heated capillary temperature was set at 200 °C to facilitate efficient complex formation. The electrospray needle voltage was set at 3.3 kV. Nitrogen was used as the sheath gas (flow 20 units) and auxiliary gas (flow 5 units) in the electrospray ionization region. The samples were introduced into the elctrospray interface by a direct infusion method using a microsyringe pump (SEG, Australia) at a flow rate of 10 mL/min. The MS/MS spectra were acquired with experimental conditions of an isolation width of 1 mass unit, an activation time of 30 msec and $q_z = 0.25$. In MS/MS mode, the parent ion molecules were manually selected one by one, and each was subjected to collision-induced dissociation (CID).

Reagents. Gly-Gly-His (99%, Sigma-Aldrich Korea), Cupric chloride dihydrate (99%, Sigma-Aldrich Korea), Nickel(II) nitrate hexahydrate (97%, Junsei chemical Co., Tokyo, Japan), Zinc nitrate hexahydrate (98%, Sigma-Aldrich Korea), Calcium chloride dihydrate (98%, Dae Jung chemical, Korea), and H₂O (HPLC grade, Merck) were used in experiments. Gly-Gly-His was dissolved in water to prepare a 2.4 \times 10⁻⁴ M solution. The four metal solutions were prepared in water at a final concentration of 2.4 \times 10⁻⁴ M. These two solutions were mixed together prior to obtaining the mass spectra.

Results and Discussion

The structural features of the [(Cu²⁺---Gly-Gly-His) – 3H⁺]⁻¹ complex in aqueous solution are shown in Figure 1.^{18,19} The [(Cu²⁺---Gly-Gly-His) – 3H⁺]⁻¹ complex is seen to possess a planar structure involving the coordination of a terminal amino nitrogen, two deprotonated amide nitrogens, and the imidazole-N3 atom. The [(Cu²⁺---Gly-Gly-His) – 3H⁺]⁻¹ planar complex between Cu²⁺ and four central nitrogen atoms (4 N) is known as the most stable structure in the four-coordination complex geometries.

Negative mode MS spectra of four metal ion complexes in aqueous solution are shown in Figure 2. The [(63 Cu²⁺, 58 Ni²⁺, 64 Zn²⁺, Ca²⁺---Gly-Gly-His) – 3H⁺]⁻¹ complexes were observed at m/z 329, m/z 324, m/z 330, m/z 306 and the [(Gly-Gly-His – H⁺)]⁻¹ peptide ion was observed at m/z 268 (Fig. 2). The most meaningful observation gleaned from the MS spectra is that the formation efficiency of [Cu²⁺, Ni²⁺---(Gly-

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Figure 1. Structure of $[(Cu^{2+}$ ---Gly-Gly-His) $-3H^+]^{-1}$ complex in aqueous solution.

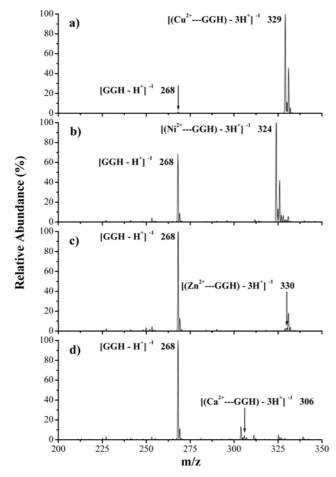


Figure 2. MS spectra in negative mode: (a) Cu^{2+} ion + Gly-Gly-His, (b) Ni^{2+} ion + Gly-Gly-His, (c) Zn^{2+} ion + Gly-Gly-His and (d) Ca^{2+} ion + Gly-Gly-His.

Gly-His $-3H^+$)]⁻¹ complex is much better than that of [Zn²⁺, Ca²⁺---(Gly-Gly-His $-3H^+$)]⁻¹ complex. The more than adequate formation efficiency of the [Cu²⁺, Ni²⁺---(Gly-Gly-His $-3H^+$)]⁻¹ complex was explained by the stabilization energy of the four-coordination planar structures in the [Cu²⁺, Ni²⁺---(Gly-Gly-His $-3H^+$)]⁻¹ complex. The reason of bad formation efficiency of the [Zn²⁺---(Gly-Gly-His $-3H^+$)]⁻¹ complex is not clear in this step. The ratios of

Table 1. The ratios of $[(Metal^{2+}--Gly-Gly-His) - 3H^+]^{-1}$ peak area to $\{[(Gly-Gly-His - H^+)]^{-1}$ peak area $+ [(Metal^{2+}---Gly-Gly-His) - 3H^+]^{-1}$ peak area $\}$ in Figure 2

	Peak Area $[(Metal^{2+}Gly-Gly-His)-3H^+]^{-1}$
	Peak Area {[(Gly-Gly-His – H) ⁺] ⁻¹ + [(Metal ²⁺ Gly-Gly-His)-3H ⁺] ⁻¹ }
Cu ²⁺	0.992
Ni^{2+}	0.689
Zn^{2+}	0.093
Ca^{2+}	0.091

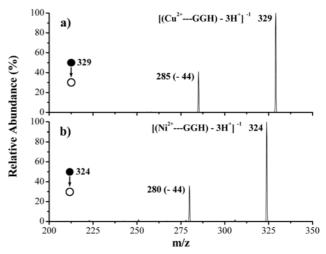


Figure 3. MS/MS spectra of $[(Cu^{2+}, Ni^{2+}--Gly-Gly-His) - 3H^+]^{-1}$ complexes: (a) $[(Cu^{2+}--Gly-Gly-His) - 3H^+]^{-1}$ complex and (b) $[(Ni^{2+}--Gly-Gly-His) - 3H^+]^{-1}$ complex.

[(Metal²⁺---Gly-Gly-His) $-3H^+$]⁻¹ peak area to {[(Gly-Gly-His $-H^+$)]⁻¹ peak area + [(Metal²⁺---Gly-Gly-His) $-3H^+$]⁻¹ peak area} are reported in Table 1. The metal isotope peak effects are also included in the area ratios. The adequate formation efficiency of the [Cu²⁺, Ni²⁺---(Gly-Gly-His $-3H^+$)]⁻¹ complex could explain why the specific Cu²⁺, Ni²⁺ transport active site of human serum albumin is similar to Gly-Gly-His peptide. ^{12,13}

The MS/MS spectra of $[(Metal^{2+}--Gly-Gly-His) - 3H^{+}]^{-1}$ complex are shown in Figure 3. The fragment ions at m/z 285 in Figure 3a and at m/z 280 in Figure 3b are thought to be a result of the common loss of a CO₂ moiety from the $[(Cu^{2+}, Ni^{2+}-Gly-Gly-His) - 3H^{+}]^{-1}$ complex at the low collision activation energy. Yang et al. reported that the fragment ion of a 44u loss corresponds to a decarboxylation from the histidine residue. ⁷ In their previous works, the CO₂loss fragment of m/z 285 was reported as the one of several fragments of the $[(Cu^{2+}-Glv-Glv-His) - 3H^{+}]^{-1}$ parent ion because of the uncontrolled collision activation energy in the anion formation MS spectrum. It is worth noting that the C- CO_2 bond of the $[(Cu^{2+}, Ni^{2+}--Gly-Gly-His) - 3H^+]^{-1}$ complex was found to be the weakest bond of the [(Cu²⁺, Ni²⁺---Gly-Gly-His) – 3H⁺]⁻¹ complex in our low energy CID-MS/ MS spectra.

The MS/MS/MS spectra of the CO₂-loss fragment that

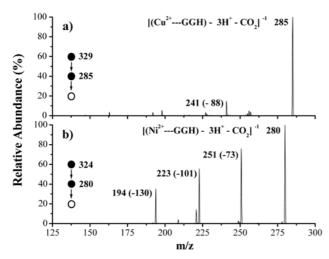


Figure 4. MS/MS/MS spectra of $[(Cu^{2+}, Ni^{2+}--Gly-Gly-His) - 3H^+ - CO_2]^{-1}$ complexes: (a) $[(Cu^{2+}--Gly-Gly-His) - 3H^+ - CO_2]^{-1}$ complex and (b) $[(Ni^{2+}--Gly-Gly-His) - 3H^+ - CO_2]^{-1}$ complex.

originated from the [(Cu²⁺, Ni²⁺---Gly-Gly-His) – 3H⁺]⁻¹ complex are shown in Figure 4. It is assumed that the observed fragments of m/z 251, m/z 223, m/z 194 in Figure 4b) are the x_2 , y_2 and x_1 ions of the $[(Ni^{2+}--Gly-Gly-His) -$ 3H⁺ – CO₂]⁻¹ complex. However, the main fragment of the $[(Cu^{2+}--Gly-Gly-His) - 3H^{+} - CO_{2}]^{-1}$ complex in a) was observed at m/z 241. The fragment of m/z 241, the ion resulting from a 44u loss from the [(Cu²⁺---Gly-Gly-His) – 3H⁺ – CO₂]⁻¹ complex, is not a fragment normally obtained in the peptide dissociation in a typical MS spectrum. The additional 44u-loss could be explained by a C₂H₄NH₂, or HCONH, or HCOCH₃ loss from the [(Cu²⁺---Gly-Gly-His) $-3H^+-CO_2$]⁻¹ complex. It is difficult to address the mechanism for the formation of these ions because of the lack of information in the collision-induced dissociation spectra. Further experimentation is needed for a better understanding of the fragmentation patterns in the [(Cu²⁺---Gly-Gly-His) – $3H^+ - CO_2$]⁻¹ MS/MS/MS spectrum.

In summary, the adequate formation efficiency of the $[Cu^{2+}, Ni^{2+}--(Gly-Gly-His - 3H^+)]^{-1}$ complex in the gas

phase MS spectra reflects what is also observed in the solution phase absorption spectra. The C-CO₂ bond is found to be the weakest bond of the $[(Cu^{2+}, Ni^{2+}--Gly-Gly-His) - 3H^+]^{-1}$ complex in our low energy CID-MS/MS spectra. The structure of the $[(Cu^{2+}, Ni^{2+}--Gly-Gly-His) - 3H^+]^{-1}$ complex in the gas phase was assumed to maintain the planar structure it held in the solution phase on the basis of the analysis of the MS and MS/MS spectra.

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