Enantioselective Synthesis of (4S,E)-4-Methylhex-2-enoic Acid and (4R,E)-4-Methylhex-2-enoic Acid

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Received January 8, 1993

(+)-(4S,E)-4-Methylhex-2-enoic acid $[(+)-1]^1$ is the key constituent of the peptide antibiotics leucinostatines possessing antibiotic, antitumoral, antibacterial and phytotoxic activities. Three syntheses have been reported² for (+)-1. In connection with our research programs to utilize optically active carbonates and sulfites as activating groups,³ we were interested in the synthesis of (+)-1. Here we report an enantioselective synthesis of (+)-1 and its enantiomer (-)-1 based on S_N2' addition of organocuprates to chiral allylic cyclic carbonates.

The acetonide 34 was prepared from (2S,3S)-2,3-O-isopropylidenedioxy-1,4-butanediol 25 in a three-step sequence via monosilylation, Swern oxidation⁶ and Wittig olefination reaction. Deprotection of the acetonide followed by carbonylation with carbonyl diimidazole afforded the allylic cyclic carbonate 4. Highly diastereoselective (>99%) S_N2' addition of 4 with MeMgBr, CuI (3 mol%), and BF₃·Et₂O afforded the allylated compound 54, which constitute the key step for the introduction of chirality. The diastereoselection was determined by NMR spectroscopy with a chiral shift reagent [1H-NMR, 300 MHZ, chiral Eu(tfc)₃]. The exclusive (E)-stereochemistry was judged by 1H-NMR (300 MHz) coupling constants of the two vinyl protons. Deprotection of the silyl group in 5 gave the diol 64, which was transformed into the target compound, (+)-1⁵, $[\alpha]_D^{25} = +47.8$ (c 0.12, CHCl₃), (lit.^{1b} $[\alpha]_D^{20} = +49.7$) by oxidative cleavage with NaIO₄ followed by NaClO₂ oxidation (Scheme 1).

Alternatively, the enantiomer (-)-1 was also synthesized

HO OSIPh₂ +Bu
$$\frac{d_1 \cdot d_2}{3}$$
 OSIPh₂ +Bu $\frac{d_2 \cdot d_3}{3}$ OSIPh₂ +Bu $\frac{d_1 \cdot d_2}{3}$ OSIPh₂ +Bu $\frac{d_2 \cdot d_3}{3}$ OSIPh₂ +Bu $\frac{d_1 \cdot d_2}{3}$ OSIPh₂ +Bu $\frac{d_2 \cdot d_3}{3}$ OSIPh₂ +Bu $\frac{d_1 \cdot d_3}{$

a) NaH, t-BuPh₂SiCl, DME, -20° C, 3 h (91%); b) (COCl)₂, DMSO, Et₃N, CH₂Cl₂, -78° C, 1 h (91%); c) n-BuLi, Ph₃P⁺CH₂-CH₂CH₃Br⁻, THF, -78° C, 10 h (63%); d) 70% AcOH, 40°C, 5 h (89%); e) CO(Im)₂, CH₂Cl₂ rt, 10 min (93%); f) MeMgBr (2 equiv), CuI (3 mol%), BF₃·Et₂O (1 equiv), THF, -78° C, 30 min (87%); g) (n-Bu)₄NF, THF, rt, 2 h (96%); h) NaIO₄, SiO₂, CH₂Cl₂, 1 h (89%); i) NaClO₂, t-BuOH, NaH₂PO₄, rt, 8 h (68%).

Scheme 1.

a) n-BuLi, $\text{Ph}_3\text{P}^+\text{CH}_2\text{CH}_3\text{Br}^-$, THF, -78°C , 10 h (75%); b) Dowex 50 W X 8 resin, MeOH, 45°C , 6 h (92%); c) CO(Im)₂, CH₂Cl₂, rt, 10 min (84%); d) EtMgBr (2 equiv), CuI (3 mol%), BF₃·Et₂O (1 equiv), THF, -78°C , 30 min (75%); e) Na, NH₃ (1), THF, -78°C , 3 h (91%); f) NaIO₄, SiO₂, CH₂Cl₂, 1 h (90%); g) NaClO₂, t-BuOH, NaH₂PO₄, rt, 8 h (67%).

Scheme 2.

from 4-O-benzyl-2,3-isopropylidene-L-threose \mathcal{T}^{5b} by the similar methodology, which is shown in Scheme 2.

Acknowledgement. Generous financial support by Korea Science and Engineering Foundation (KOSEF)-the Organic chemistry Research Center (OCRC) is gratefully acknowledged.

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- 4. Satisfactory spectral and physical data were obtained for all new compound and are in accord with the assigned structure. Selected spectral data are as follows. (+)-1: 1 H-NMR (300 MHz, CDCl₃) δ 0.92 (t, 3H), 1.05 (d, 3H), 1.45 (m, 2H), 2.22 (m, 1H), 5.80 (d, 1H, J=16 Hz), 6.89 (dd, 1H, J=16.8 Hz), 12.25 (s, 1H). IR (neat) 3600-2400, 1685, 1640 cm⁻¹. [α] $_{D}^{25}$ = +47.8 (c 0.12, CHCl₃) (-)-1: [α] $_{D}^{25}$ = -47.2 (c 0.14, CHCl₃) 5: TLC; SiO₂, EtOAc/hexane 1:3, R_f =0.71. 1 H-NMR (200 MHz, CDCl₃) δ 0.82 (t, 3H, J=7.5 Hz), 0.96 (d, 3H, J=6.9 Hz), 1.08 (s, 9H), 1.26 (m, 2H), 2.05 (m, 1H), 3.56 (m, 1H), 3.65 (m, 1H), 4.20 (m, 1H), 5.35 (dd, 1H, J=15.5, 6.5 Hz), 5.62 (dd, 1H, J=15.5,

7.5 Hz), 7.38-7.46 (m, 6H), 7.67-7.70 (m, 4H). IR (neat) 3400, 3050, 2950 cm⁻¹, $\lceil \alpha \rceil_D^{25} = +8.0$ (c 0.15, CHCl₃). MS (m/e) 325 (M-tBu), 269, 247, 199 (base peak), 181, 139, 135, 109, 57. 6: ¹H-NMR (300 MHz, CDCl₃) δ 0.85 (t, 3H), 0.95 (d, 2H), 1.25-1.34 (m, 6H), 2.05 (m, 1H), 3.50 (m, 1H), 3.65 (m, 1H), 4.22 (m, 1H), 5.40 (dd, 1H), 5.65 (m, 1H). IR (neat) 3300, 2950 cm⁻¹, $\lceil \alpha \rceil_D^{25} = +1.82$ (c 0.17, CHCl₃). 10: TLC; SiO₂, EtOAc/hexane 1:5, $R_f = 0.33$, ¹H-NMR (300 MHz, CDCl₃) δ 0.86 (t, 3H, J = 7.5 Hz), 0.97 (d, 3H, J = 6.9 Hz), 1.27-1.37(m, 1H), 3.57 (dd, 1H, J = 11.4, 7.8 Hz), 3.69 (dd, 1H, J = 10, 3.6 Hz), 4.22 (m, 1H), 4.58 (s, 2H), 5.38 (ddd, 1H, J = 15.5, 6.6, 1 Hz), 5.65 (ddd, 1H, J = 15.5, 6.6, 1 Hz), 7.35 (s, 5H). 11: $\lceil \alpha \rceil_D^{24} = -39.8$ (c 3.0, CHCl₃).

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Synthesis of Steroidal Cyclophosphamide, 2-Bis (2-chloroethyl)amino-2-oxo-6-(5α -cholestanyl)-1, 3,2-oxazaphosphorinane

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Received January 8, 1993

Cyclophosphamide and its analogues are important clinical agents in the treatment of cancer.1 We have prepared steroidal cyclophosphamides (1a and 1b). The approach used for the synthesis of 1a and 1b is outlined in Scheme 1. Treatment of cholestanone (2) with n-butyllithium and acetonitrile gave a 72.5% yield of β-hydroxynitrile derivative 3², which was subsequently reacted with LiAlH4 to give aminoethyl derivative 4.3 Cyclization of 4 with bis(2-chloroethyl)phosphoramidic dichloride (5) in the presence of 2 equiv. of Et₃N afforded crude mixtures of 1a and 1b, which were chromatographed on silica gel with EtOAc: CH₂Cl₂: hexane=2:2:1 to give analytically pure crystals of the faster (mp. 192-194°C) and slower (mp. 178-180°C) eluting diastereomers of 1a and 1b in 58% yield. Assignment of cyclophosphamide structures to the faster and slower eluting diastereomeric cyclization products has been suggested by the IR, ¹H-NMR, ³¹P-NMR⁴, and ¹³C-NMR.

Our measurements of 1a and 1b indicated the ¹H-NMR chemical-shift difference between the NH resonances at 2.73 and 2.50 ppm for the faster and slower eluting diastereomers of 1a and 1b, respectively. The substantial deshielding (0.23 ppm) of N-H proton thus exhibited by the faster moving

compound 1a, suggests more efficient intramolecular H-bonding to the adjacent P=O functionality. This difference in H-bonding was also founded in ¹³C-NMR by the deshielding of chemical shift[41.9 ppm (-NH-CH₂-)] in the proposed 1a, as opposed by the shielding of chemical shift [36.0 ppm (-NH-CH₂-)] in the proposed 1b. These compounds may have a greater impact as anticancer agents by their lipophilicity. Compounds 1a and 1b were found no activity against Hepatoma cells⁵.

Experimental

3-Cyanomethyl-5α-cholestan-3-ol (3). To a stirred solution of 1.6 M n-butyllithium in 9.5 ml (15 mmol) hexane, at -80°C under nitrogen, was rapidly added a solution of 0.82 ml (15 mmol) of acetonitrile in 30 ml of anhydrous THF. After stirring for 1 hr, the resulting white suspension was treated with a solution of 3.0 g (7.5 mmol) 2 in 10 ml of THF. The cold-ice bath was removed and stirred for additional 10 min before it was poured into ice-water hydrochloric acid. The aqueous layer was extracted with three 50 ml portions of Et₂O. The combined ether extracts were dried(MgSO₄) and evaporated in vaccuo, and the residual crude product was chromatographed on silica gel with CH₂Cl₂ as an eluent, and obtained 2.4 g (73% yield) of white solids, mp. 158-159 °C: 1H-NMR (CDCl₃) 8 2.6 (s. 2H, -CH₂CN), 0.6-2.0 (m, H steroid); IR (KBr) 3480 (-OH), 2930, 2255 (-CN), 1460, 1370, 1080, 1050 cm⁻¹.

3β-Aminoethylene-5α-cholestan-3-ol (4). To a stirred solution of 1.7 g (3.9 mmol) of 3 in 150 ml of anhydrous THF was added in small portions, 0.75 g (19.5 mmol) of lithium aluminum hydride. The mixture was refluxed with stirring for 17 hrs. After decomposing excess lithium aluminum hydride with 0.75 ml water and 2.3 ml of 20% NaOH, the mixture was filtered and filtrate was evaporated in vaccuo to obtain yellow oily residues (45% yield). All attempts

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